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In Silico Post Translational Analysis of Functional Single Nucleotide Alterations in Human TERT Gene Associated with Acute Myeloid Leukemia

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INTRODUCTION

Acute myeloid leukemia (AML) is a heterogeneous hematologic malignancy of the bone marrow in which hematopoietic precursors maturation is seized in the early stages of development [1]. A clonal disease characterized by the piling up of somatic acquired genetic mutations in hematopoietic progenitor cells that modify the normal mechanisms of regeneration, proliferation and differentiation [2]. The most prevalent acute leukemia in adults is AML, and as people get older, its extent rises [3]. It remains a fatal disease with a less than 30% 5-year survival rate [4]. AML has variable symptoms, being presented clinically as a combination of cytopenia, which includes weakness, fever, abdominal pain, pallor, shortness of breath, fatigue, easy bruising and bleeding with an elevated

ABSTRACT

Acute myeloid leukemia (AML) refers to a diverse assemblage of hematological malignancies that constitute clonal expansion of immature myeloid progenitor cells in the peripheral blood and bone marrow. TERT gene ensures telomeres maintenance, chromosome stability and prevention of malignancy. The TERT gene has several single nucleotide polymorphisms (SNPs) that have been linked to a number of diseases, including AML. Objective: To classify the harmful TERT gene mutations, and to analyze them using various computational approaches at structural, functional and translational expression levels Methods: National Centre for Biotechnology Information (NCBI) database was used to retrieve nsSNPs of TERT gene (Q53H, V170M, A184T, S255Y, A288V, H412Y, I540M, R631W) reported in AML and they were analyzed using various bioinformatics tools. Results: In this in silico analysis, it was observed that seven out of eight SNPs had a damaging effect; they could affect the protein stability, protein-protein interactions, hydrophobicity, protein folding, three-dimensional structure, secondary structure and conservation profile. 3D models were generated and validated by various tools and the structural effect of these alterations was observed on protein function that was destabilizing to the RNA folding, protein-protein interactions and other functionally associated proteins. Analysis of post translational modifications showed no significant effect of these mutations. Conclusions: These SNPs could be used in future as potential targets in disease diagnosis, biological markers and protein studies.

infection risk, weight loss, nausea, vomiting and dysphagia [5]. A large and diverse group of genetic and environmental variables have been proposed [6]. The primary cause of AML is thought to be acquired genetic anomalies [7]. In humans at chromosome 5p15.33 TERT gene is located which encodes telomerase reverse transcriptase. The 1132 amino acid polypeptide produced by it is translated into a 130 kD active TERT protein [8]. TERT is an important part and catalytic subunit of the telomerase holoenzyme [9]. The 42 kb long TERT gene contains 15 introns, 16 exons, and a promoter core of 260bp [10]. It has a vital role in the maintenance of telomeres, chromosome stability and preventing malignancy [11]. The Catalytic component's (TERT) expression of telomerase triggers its reactivation

during carcinogenesis in the majority of human malignancies. This may occur by means of both methylation and mutations at TERT promoter (TERT*p*)[12]. TERT mutations, which occur commonly (2- 19%) in bone marrow failure syndromes, are linked to an elevated risk of MDS/AML [13]. TERT gene amplification is highly related to hematological malignancies, with a greater prevalence in AML patients (53.3%) [14]. Bioinformatics tools are time saving and cost effective [15]. This study focuses on the thorough *in silico* analysis that pinpoint and examine the most pathogenic mutations of the TERT gene.

METHODS

The comprehensive in silico analysis was performed by using different softwares. Human TERT gene's data and TERT protein sequence was retrieved from NCBI (https://www.ncbi.nlm.nih.gov/) and UniProtKB (http://www.uniprot.org/uniprot/) respectively. SNPs found in TERT gene were obtained from dbSNP database (http://www.ncbi.nlm.nih.gov/SNP/). To estimate the effect of amino acid alterations on the pathogenicity and the functionality of protein FATHMM (Functional Analysis Through Hidden Markov Models) http://fathmm. biocompute.org.uk [16], PolyPhen-2 (Polymorphism phenotyping v2)(http://genetics.bwh.harvard.edu/ggi/cgibin/ggi2.cgi) [17], and SIFT (sorting intolerant from tolerant) algorithm accessed via https://sift.bii.astar.edu.sg/[18] were used which showed that whether the mutation is damaging or benign. Mutation Cutoff Scanning Matrix (mCSM) (http://biosig.unimelb.edu.au/mcsm/ stability)[19] and MUpro(http://mupro.proteomics.ics.uci. edu/) [20] were used for protein stability analysis which showed the results in the form of $\Delta\Delta G$ values. For the conservation analysis Consurf (http://consurf.tau.ac.il) was used it assigned the scores ranging from 1-4 being variable, 5-6 being average and 7-9 being conserved [21]. Project HOPE (Have (y) Our Protein Explained) available at "https://www3.cmbi.umcn.nl/hope/" was used to evaluate the structural and biochemical effects of single point mutations [22]. SWISS-MODEL (http://swissmodel. expasy.org/) online web service was used for homology modelling of wild and mutant types of TERT protein [23]. 3D structures of proteins were generated and quality evaluation of the generated models was performed by some parameters like GMQE, QMEAN Z-score. SOPMA (Self-Optimized Prediction Method with Alignment) tool (https://npsa- prabi.ibcp.fr/cgibin/npsa_automat. pl?page=/ NPSA/npsa_sopma.html) was used for the secondary structure analysis. It gives the outcome as percentage composition of α - helix, β -sheet, turns, and random coil [24]. The online version of RNAfold Web Server based on the Vienna RNA package available at (http://rna.tbi.univie.ac.at/cgi- bin/RNAWebSuite/ RNAfold.cgi) [25] was used to detect single nucleotide alterations' influence on secondary structure of RNA. For the prediction of post translation modifications and mRNA expression cBioPortal (http://www.cbioportal.org/) [26], UALCAN portal (http://ualcan.path.uab.edu/analysisprot.html) [27], GEPIA (Gene Expression Profiling Interactive Analysis), a web-based tool (http:// gepia.cancer-pku.cn/)[28], Gene Set Enrichment Analysis (GSEA)[29] and Cytoscape an open-source software for integration, visualization and analysis of biological networks association analysis were used[30].

RESULTS

Missense SNPs of the TERT include (Q53H, V170M, A184T, S255Y, A288V, H412Y, I540M and R631W) were retrieved from the NCBI. It was predicted by Fathmm that all the SNPs were damaging. SIFT showed that 4 out of 8 SNPs were damaging and only one mutation (A184T) predicted benign by Polyphen-2(Table 1).

Table 1: Functional analysis of mutations in the TERT gene by using in silico programs

SNP Ids	SNPs	SIFT	Fathmm	Polyphen-2
rs1060503006	Q53H	Not tolerated	Damaging	Damaging
rs387907248	V170M	Not tolerated Damaging		Damaging
rs773758089	A184T	Tolerated	Damaging	Benign
rs1751207450	S255Y	Tolerated	Damaging	Possibly Damaging
rs774657340	A288V	Tolerated	Damaging	Possibly Damaging
rs34094720	H412Y	Tolerated	Damaging	Damaging
rs797046041	1540M	Not tolerated	Damaging	Damaging
rs1194223999	R631W	Not tolerated	Damaging	Damaging

mCSM and MUpro were employed to examine the impact of these SNPs on protein stability. It was predicted by both the tools that 7 out of 8 mutations were decreasing the protein stability except H412Y (Table 2).

Table 2: Change in protein structural stability of TERT gene by

 single point mutations estimated through mCSM and Mupro

	m	CSM	Mupro		
SNPs	∆∆G (kcal/mol)	Stability	∆∆G (kcal/mol)	Stability	
Q53H	-0.747	Destabilizing	-1.670	Decrease	
V170M	-0.107	Destabilizing	-0.439	Decrease	
A184T	-0.512	Destabilizing	-0.465	Decrease	
S255Y	-0.743	Destabilizing	-0.497	Decrease	
A288V	-0.325	Destabilizing	-0.730	Decrease	
H412Y	0.471	Stabilizing	0.0266	Increase	
1540M	-0.767	Destabilizing	-0.659	Decrease	
R631W	0.188	Stabilizing	-0.393	Decrease	

ConSurf predicted the conservation profile of the SNPs. According to the output of the ConSurf web server, it was predicted that 2 out of 8 variants (V170M, R631W) were conserved residues with a conservation score range of 7-9

shown in Table 3 and Figure 1.

Table 3: Analysis of evolutionary conservation profile of SNPs inTERT gene by ConSurf

SNPs	Conservation score	Conservation scale status
Q53H	5	Average
V170M	7	Conserved
A184T	3	Variable
S255Y	3	Variable
A288V	2	Variable
H412Y	5	Average
I540M	5	Average
R631W	9	Conserved

MPRAPRCRAV	RSLIDESHARE	VLPPATEVRR	LGPOGWRINO	RGEPAAFRAL
eeeeebebb	eebbeeebee	bbebeebbee	becebeebbe	eeeeebbebb
			-	e
51 53	61	71	81	91
NAOC LUCO PW	DARPPPAAP	PROVICERIA	VARVLORLON	RGARNVLAFC
3 66 8	e e	f a ffa		f foo o
101	111	121	131	141
ALLDGARGG	PPEAFTTSVR	SYLPHTVIDA	LRGEGANGLL	LRRVGDDVLV
bbbbeeeee	eeeebebebe	Cobcochece	bebeebbebb	beebeeebbb
151	161 170	171	181 184	1.91
HLLARCALFY	LVAPSCAYCV	COPPLYOLGA	ATCARPPPHA	SCPRREGCE
bbbeebbbbb	bbbeebbbeb	beeebeeebe		
201	211	221	231	241
RAMANDVMEA	GVPLGLPAPG	ARRAGE		
251 255	261	271	281 288	291
VGCGSTANDG	RTRGPSDRGP	CVVSPARPAE	EATSLEGALS	GTRESEPSVG
		eebeeebee		
301	311	321	331	341
ROHHAGPPST	SIPPIPWDTP	O P P V Y A E T K H	FLYSSGDKEQ	LRPSLLSSL
		b b	b b b b e e e e e	b = = b b b b = = =
BPSLTGABB	YETIGEGER	MPGTPBBEP	BLPOTT WOME	PLELECENT
eeeeebeeb	bbbbbbeeee		obccobbobb	ebbeebbeee
-	412			
401	411	421	431	441
AQCPEQVLEK	THEPLRAAVT	PAAGVCARER	POGEVAAPE	EDTDPRRLVQ
451	461	471	401	491
LLRQHSSPWQ	VXGFVRACLR	RLVPPGLWGS	RHNERRELRN	TKKFISLGKH
bbeeseebe	bbbbbbeebbe	obbeeebbbe	eeeeebbee	beebbeebee
501	511	521	531 54	0 541
AKLELTW	KMSVRDCAWL	RRSPGVGCVP	ARHRLREEI	AKFLHWLMS
••b•b•bbb	• b • b • • b • b b	occoccbe	e e e e e e e e e e e	b b e b b b b b b e
VYEVELBSE	EXVIETTEOR	NRLFTYRESY	SALOSIGIB	OLLKBYOLBE
b b b b e b b e b b	bebeeeeee	o e b e b b e e e b	beebeebebe	ocbocbobce
		£ • £		
601	611	621	631	641
LBEAEVROHR	EARPALLTER	LRFIPKPDGL	RPIVNMDYVV	GARTERREKR
f	bebebe	afa ff fa	fas	
The conserve	ation scale:			
? 1 2 3 4	5 6 7 8 9			

Variable Average Conserved

An exposed residue according to the neural network algorithm.
 A buried residue according to the neural network algorithm.

Figure 1: Representation of conservation profile of TERT gene's amino acids in the form of various colors using Consurf.

HOPE was used to examine the structural as well as functional impacts of single amino acid changes on protein. It claimed that all the mutant residues were bigger in size than the wild residue. Only one mutation R631W altered the charge of amino acid from positive to neutral and this difference in charge will disturb the ionic interaction made by the original, wild-type residue. The remaining 7 SNPs were predicted not to affect the charge. As interpreted by HOPE 2 mutant residues (H412Y and R631W) were more hydrophobic than wild-type.

Table 4: Evaluation of amino acid replacement's effect on the structure of TERT protein with reference to wild residue by HOPE

SNPs	Size	Change of Charge	Hydrophobicity
Q53H	W <m< td=""><td>Not affected</td><td>Not affected</td></m<>	Not affected	Not affected
V170M	W <m< td=""><td>Not affected</td><td>Not affected</td></m<>	Not affected	Not affected
A184T	W <m< td=""><td>Not affected</td><td>Wildtype residue (A) is more hydrophobic</td></m<>	Not affected	Wildtype residue (A) is more hydrophobic

S255Y	W <m< td=""><td>Not affected</td><td>Not affected</td></m<>	Not affected	Not affected
A288V	W <m< td=""><td>Not affected</td><td>Not affected</td></m<>	Not affected	Not affected
H412Y	W <m< td=""><td>Not affected</td><td>Mutant residue(Y) is more hydrophobic</td></m<>	Not affected	Mutant residue(Y) is more hydrophobic
1540M	W <m< td=""><td>Not affected</td><td>Not affected</td></m<>	Not affected	Not affected
R631W	W <m< td=""><td>+ve to neutral</td><td>Mutant residue (W) is more hydrophobic</td></m<>	+ve to neutral	Mutant residue (W) is more hydrophobic



Figure 2: The native amino acid (left) and the mutant amino acid (right) are depicted in schematic form via HOPE

SWISS-MODEL was used to generate the TERT protein homology model. The QMEAN-Z score (-5-0) and GMQE values (0-1) revealed that there was greater compatibility between template and target structure of similar size and the alignment was quite precise (Table 5 and Figure 3).

SNPs	Template Query No.	Sequence Identity (%)	GMQE	OMEAN Z-score
Wildtype	7trd.1. B	100	0.72	-2.62
Q53H	7trd.1. B	99.91	0.72	-2.78
V170M	7trd.1. B	99.91	0.72	-2.65
A184T	7trd.1. B	99.91	0.72	-2.77
S255Y	7trd.1. B	99.91	0.72	-2.43
A288V	7trd.1. B	99.91	0.72	-2.24
H412Y	7trd.1. B	99.91	0.72	-2.67
1540M	7trd.1. B	99.91	0.72	-2.90
R631W	7trd.1. B	99.91	0.72	-2.86





Figure 3: Photographs of Protein structure using Swiss Model. (A) Wildtype(B)Q53H(C)V170M(D)A184T(E)S255Y(F)A288V(G)H412Y (H)I540M(I)R631W

SOPMA was used for the secondary structure analysis and outcome was in the form of percentages of different parameters which are shown in table 6 and figure 4.

Table 6: Prediction of Secondary Structure of TERT protein using

 SOPMA

SNPs	Alpha Helix (%)	Extended Strand (%)	Beta Turn (%)	Random Coil (%)
Wildtype	44.08	10.51	3.71	41.70
Q53H	43.37	10.78	4.06	41.78
V170M	43.20	11.22	3.80	41.78
A184T	42.76	10.95	4.15	42.14
S255Y	42.40	10.95	3.98	42.67
A288V	42.31	11.31	3.71	42.67
H412Y	42.84	11.04	3.80	42.31
1540M	42.93	11.31	3.89	41.87
R631W	42.58	11.22	3.71	42.49

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	10	20	30	40	50	60	70
	I	I	l.	I	<u>n</u>	1	
1PRA	PRCRAVRSLL	RSHYREVLPLA	ATEVRRLGPQ	GWRLVQRGD	PAAFRALVAQCL	VCVPWDARP	PPAAPS
RON	CINELVARY		MARCEALL	DCARCODE	Chrinninninne		CALICLE
RQU	SCERELVARV	LURLCERGARN	IVLAFGFALL	DGARGGPPE	AFTISVRSYLPN	hebbbbbbbb	GAWGEL
PPL			CANCERE	VOLGAATO		PLOCEPAWA	
hh	chhhhhhhhhhh	hbbbbeeectt	contractor	CTQCGAATQ		coccoch	hbbbbbt
VPL	GLPAPGARR	GGSASRSLPLE	KRPRRGAAF	PEPERTPVGO	SWAHPGRTRGF	SDRGECVVS	PARPAE
ccc	ccccccccc	ccccccccc	ecceccec	cccccccc		cccceeec	cccchh
ATS	LECALSGTRH	SHPSVGRQHHA	GPPSTSRPP	RPWDTPCPP	VYAETKHELYSS	GDKEQLRPS	FLLSSL
hhh	heccecce			ceccecee	eecccceeeecc	ccccccch	hhhhhc
RPSL	TGARRLVETI	FLGSRPWMPGT	PRRLPRLPQ	RYWQMRPLF	LELLGNHAQCPY	GVLLKTHEF	LRAAVT
ccc	ccchhhhhee	eeccccccc	cececcel	hhhhhhhhh	hhhhh <mark>ccccc</mark> ł	hhhhhtoco	ccccc
PAAG	SVCAREKPQGS	VAAPEEEDTD	PRRLVQLLRQ	HSSPWQVYG	FVRACLRRLVPF	GLWGSRHNE	RRFLRN
ccc	ecceccecce	cecececee	cchhhhhh	hcccchhhhh	hhhhhhhccco	cececee	hhhhhh
TKKI	ISLGKHAKLS	LQELTWKMSVE	RDCAWLRRSF	GVGCVPAAE	HRLREEILAKFI	HWLMSVYV	/ELLRSF
hhł	neetttcchhh	hhhhhhh <mark>cc</mark>	ccceeccc	cccccchh	hhhhhhhhhhhh	հերեթեր	hhhhtt
FYV	TETTFQKNRLF	FYRKSVWSKL	QSIGIRQHL	RVQLRELSE	AEVRQHREARP/	ALLTSRLRFI	PKPDGL
eeee	ecccccccee	eechhhhhhh	hhhcchhh	hhhhhhcch	hhhhhhhhcco	cccheeee	CCCCCC
(PI)	MMDYVVGAR	FREEKRAERL	ISKVKALFSV	LNYERARRP	GLLGASVLGLDL	THRAWKIE	LEVKAQ
DDD		CAVDTIDODRI	TEVTACTT	DONTYCVPP			TITDLO
SPPI	ceeeeeecht	hhhcccchhh	hhhhhhhhh	ttchbbbbb	heeeeccttch	hhhhhhhh	chhhcc
PYME	ROEVAHLOETS	PLEDAWYTEOS	SSLNEASS	SLEDVEL REM	CHHAVRIEGKSY	WOCOGTPOO	STIST
hhh	hhhhhhhhhh	hhhheeeehh	hcccchh	hhhhhhhhh	hhhheeetccee	eecccccc	cehhhh
CSI	CYGDMENKLE	AGIRRDGLLLF	REVDDELLVT	PHLTHAKTE	LRTLVRGVPEY		WNEPVE
hhh	hhhhhhhhhh	hhcccttheed	hhhheee	ccchhhhhh	hhhhttccccd	eeecttcce	eeeecc
DEAL	GGTAFVQMPA	HGLFPWCGLL	DTRTLEVQS	DYSSYARTS	IRASLTENRGE	AGRNMRRKL	FGVLRL
ccc	ecceccecc	ccccccceee	eccceeeee	hhhhhcchh	hhhheeeeccco	tthhhhhhh	hhhhht
CHS	LELDLQVNSL	QTVCTNIYKI	LLQAYRFH/	CVLQLPFHQ	QVWKNPTFFLR\	ISDTASLC)	SILKAK
teed	eeeeccccch	hhhhhhhhhh	hhhhhhhh	hhhhccccc	cccccchhhhh	hhhhhhhh	hhhhcc
AGN	ISLGAKGAAGF	PLPSEAVQWLCH	QAFLLKLT	RHRVTYVPLL	GSLRTAQTQLSP	RKLPGTTLTA	LEAAAN
ccc	eeccccccc		hhhhhhh	ccchhhhhh	hhhhhhhhhhh	h <mark>cc</mark> hhhhh	hhhhcc
PALF	SDFKTILD						
ccc	cchhhhhh						

Sequence length :	1132						
SOPMA :							
Alpha helix	(Hh)	:	499	1s	44.08%		
3 ₁₀ helix	(Gg)	:	ø	is	0.00%		
Pi helix	(Ii)	I	0	is	0.00%		
Beta bridge	(Bb)	:	0	is	0.00%		
Extended strand	(Ee)	:	119	is	10.51%		
Beta turn	(Tt)	ī	42	is	3.71%		
Bend region	(Ss)	z	0	is	0.00%		
Random coil	(Cc)	:	472	15	41.70%		
Ambiguous state	s (?)	:	6	a is	0.00%		
Other states		:	ø	is	0.00%		
			~			(TEDT	

Figure 4: Representing Secondary Structure of TERT protein predicted by SOPMA

The effect of gene mutations on the secondary structure of RNA was examined using the RNAfold tool of the Vienna package. Every mutation resulted in inappropriate RNA folding compared to the wild type, which affects mRNA localization and protein translation (Figure 5).





Figure 5: TERT gene's Mutations effect on RNA secondary structure shown by RNA fold server. (A) Wildtype (B) 053H (C) V170M(D)A184T(E)S255Y(F)A288V(G)H412Y(H)I540M(I)R631W The secondary structure of RNA with a higher MFE value was one that was more stable(table 7).

|--|

SNPs	Minimum Free Energy (kcal/mol)
Wildtype	-797.77
Q53H	-804.47
V170M	-712.70
A184T	-826.07
S255Y	-832.67
A288V	-789.67
H412Y	-788.08
1540M	-797.97
R631W	-795.87



Figure 6: Representation of missense mutations and PTM sites by cBioPortal

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However, PTM sites were evaluated in which phosphorylation was the main type of PTM in TERT, with a total of 13 sites, followed by acetylation which had only 1 site. These phosphorylation sites were in the Telomerase-RBD and RVT-1 domain, thus might affect the function of protein (Figure 6). To analyze the expression and clinical significance of TERT gene in AML, UALCAN was used. The level of methylation of TERT gene in the promoter region can lead to the development of AML. TERT promotor was significantly hyper- methylated in both male and female (Figure 7 (A)). Whereas, in individuals with age group 21 years to 100 years were TERT promotor was significantly hypermethylated as shown in Figure 7(B).

Promoter methylation level of TERT in LAML =









GEPIA was used to obtain Transcripts per million (TPM) which showed that in tumor tissues T (n=170), the TERT gene's mRNA expression levels were noticeably greater as compared to in normal tissues N(n=70) and were shown as red and green dot plots respectively. To investigate the association between TERT mRNA expression and patient

prognosis in AML, overall survival rates were obtained using the Cox regression model which determines the relationship between variables and survival rates. TERT mRNA expression levels were visualized using Kaplan-Meier survival curves which showed no significant correlation between TERT expression and percent survival in AML (Figure 8 A and B).



Figure 8: (A) TERT mRNA expression in LAML (B) Overall survival comparison between low and high TERT groups in AML shown by GEPIA

Different biological functional gene sets were analyzed by the GSEA technique to determine the effect of TERT mutations on the protein functionality and different pathways. The GSEA results are shown as enrichment plot in the form of enrichment score. The highest divergence from zero experienced during a random walk is known as the enrichment score (ES) and is plotted against the y-axis in the graph. The biological processes involved in the positive regulation of E2F targets, G2M-Checkpoints, mitotic spindle, and MYC_Target_V1 were considerably enriched in the GSEA analysis of enrichment. This implies that TERT mutations in AML patients may influence transcription, cell growth, apoptosis, and cell adhesion, which may affect the development of the disease and prognosis. All these enrichment plots were positively correlated(Figure 9A-D).





Figure 9: GSEA representing Enrichment Plots (A) E2F Targets Hallmark (B) G2M checkpoint Hallmarks (C) Hallmark mitotic spindle(D)Hallmark MYC Targets V1

Cytoscape was used to generate a network of association of TERT protein with other related proteins which are involved in AML. A hub of 30 genes was generated which showed the direct link of TERT with 4 other genes and indirect association with many (Figure 10).



Figure 10: TERT association with other genes in AML via Cytoscape

DISCUSSION

Single nucleotide polymorphisms (SNPs) are single-base alterations that have a role in the pathophysiology of various ailments as well as variation in human biology. Previous research revealed that using experimental methodologies to predict effects of nsSNPs on the structure and functionality might be time-consuming and expensive. Computer simulations (in silico analysis) have recently emerged as an excellent method for comprehending disease-related mutations and their consequences in protein structures [31, 32]. TERT gene was selected for the in-silico analysis, as previously it has not been carried out. All the SNPs (053H, V170M, A184T, S255Y, A288V, H412Y, I540M, R631W) were retrieved from the dbSNP of the NCBI database. The functional analysis was done by SIFT, Fathmm and PolyPhen-2 were employed for the functional analysis which showed all the SNPs were damaging except A184T. The coding region of the TERT

gene is essential for accurate telomerase activity and telomere length maintenance [33], the functional effect of these coding region mutations leads to overexpression of TERT and abrupt telomerase activity and TL that causes abnormal proliferation and becomes a cause of cancer. Stability analysis showed all mutations were destabilizing to protein structure except one H412Y. Most often, decreased stability is the cause of protein functionality loss brought on by mutations [34]. PROJECT HOPE analyzed that the charge, size, and hydrophobicity values of wild-type residues and mutant residues were different. Two mutations were falling in the conserved region as predicted by conSurf. Previous study by Shaw suggest that evolutionarily conserved regions are the potential sites for disease causing point mutations [35]. SWISS-MODEL quality assessment parameter suggested the accuracy of generated models. SOPMA showed most mutations were in the coiled region and few were in the helical region. Vienna package showed abnormal RNA folding of mutant residues from the wild type due to single base alteration. Covalent modifications of the polypeptides after their synthesis to make them functional are called post translation modifications (PTMs) [36]. Finding disease- associated nsSNPs changing PTM sites can help to assess the various PTM candidates involved in diseases [37]. This study showed the detail analysis of missense substitutions on PTMs. cBioPortal predicted that none of the most deleterious nsSNPs positions correlated with a potential PTM sites. It could be estimate that most of them were the silent mutations. UALCAN showed the promotor methylation level of TERT gene. Hyper-methylation at THOR (TERT hyper- methylated oncological region) was often observed in AML [38]. In malignant tissues expression levels of TERT's mRNA were higher according to GEPIA. Enrichment plot was obtained by GSEA which showed that different hallmarks have an extremely overrepresented TERT gene. Cytoscape showed the association hub of TERT with other proteins involved in AML. Mutations may influence prognosis and contribute to the development of the disease by affecting transcription, apoptosis, cell adhesion as well as cell division in patients of AML.

CONCLUSIONS

This *in silico* analysis of the TERT's functional SNPs offered a substantial understanding of their damaging effects. TERT gene enhanced expression will occur, leading to the development of cancer and an adverse prognosis of AML.

Authors Contribution

Conceptualization: AM Methodology: AMA

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Formal analysis: AT, KJ Writing-review and editing: AMA, AM

All authors have read and agreed to the published version of the manuscript.

Conflicts of Interest

The authors declare no conflict of interest.

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REFERENCES

- [1] Obeagu El and Babar Q. Acute Myeloid Leukaemia (AML): The Good, the Bad, and the Ugly. International Journal of Current Research and Medical Sciences. 2021; 7(7): 29-41.
- [2] Bullinger L, Döhner K, Döhner H. Genomics of acute myeloid leukemia diagnosis and pathways. Journal of Clinical Oncology. 2017 Mar; 35(9): 934-46. doi: 10.1200/JCO.2016.71.2208.
- [3] Irish W, Ryan M, Gache L, Gunnarsson C, Bell T, Shapiro M. Acute myeloid leukemia: a retrospective claims analysis of resource utilization and expenditures for newly diagnosed patients from first-line induction to remission and relapse. Current Medical Research and Opinion. 2017 Mar; 33(3): 519-27. doi: 10.1080/03007995.2016.1267615.
- [4] Levin M, Stark M, Ofran Y, Assaraf YG. Deciphering molecular mechanisms underlying chemoresistance in relapsed AML patients: Towards precision medicine overcoming drug resistance. Cancer Cell International. 2021 Dec; 21(1): 1-6. doi: 10.1186/s12935-021-01746-w.
- [5] Kabel A, Zamzami F, Al-Talhi M, Al-Dwila K, Hamza R. Acute myeloid leukemia: A focus on risk factors, clinical presentation, diagnosis and possible lines of management. Cancer Research Treatment. 2017; 5: 62-7.
- [6] Tebbi CK. Etiology of acute leukemia: A review. Cancers. 2021 May; 13(9): 2256. doi: 10.3390/ cancers13092256.
- [7] Rehman A, Akram AM, Chaudhary A, Sheikh N, Hussain Z, Alsanie WF, et al. RUNX1 mutation and elevated FLT3 gene expression cooperates to induce inferior prognosis in cytogenetically normal acute myeloid leukemia patients. Saudi Journal of Biological Sciences. 2021 Sep; 28(9): 4845-51. doi: 10.1016/j.sjbs.2021.07.012.
- [8] Ly H. Telomere dynamics in induced pluripotent stem cells: potentials for human disease modeling. World Journal of Stem Cells. 2011 Oct; 3(10): 89. doi: 10.4252/wjsc.v3.i10.89.

- [9] Dratwa M, Wysoczańska B, Łacina P, Kubik T, Bogunia-Kubik K. TERT-regulation and roles in cancer formation. Frontiers in Immunology. 2020 Nov; 11: 589929. doi: 10.3389/fimmu.2020.589929.
- [10] Akincilar SC, Unal B, Tergaonkar V. Reactivation of telomerase in cancer. Cellular and Molecular Life Sciences. 2016 Apr; 73: 1659-70. doi: 10.1007/s00018-016-2146-9.
- [11] Ding D, Zhou J, Wang M, Cong YS. Implications of telomere-independent activities of telomerase reverse transcriptase in human cancer. The FEBS Journal. 2013 Jul; 280(14): 3205-11. doi: 10.1111/ febs.12258.
- [12] Barthel FP, Wei W, Tang M, Martinez-Ledesma E, Hu X, Amin SB, et al. Systematic analysis of telomere length and somatic alterations in 31 cancer types. Nature Genetics. 2017 Mar; 49(3): 349-57. doi: 10.1038/ng.3781.
- [13] Du HY, Pumbo E, Ivanovich J, An P, Maziarz RT, Reiss UM, et al. TERC and TERT gene mutations in patients with bone marrow failure and the significance of telomere length measurements. Blood, The Journal of the American Society of Hematology. 2009 Jan; 113(2): 309-16. doi: 10.1182/blood-2008-07-166421.
- [14] Abdelrahman AH, Eid MM, Hassan M, Eid OM, AbdelKader RM, AlAzhary NM, et al. Telomerase reverse transcriptase gene amplification in hematological malignancies. Egyptian Journal of Medical Human Genetics. 2019 Dec; 20: 1-9. doi: 10.1186/s43042-019-0036-z.
- [15] Dana H, Mahmoodi Chalbatani G, Gharagouzloo E, Miri SR, Memari F, Rasoolzadeh R, et al. In silico analysis, molecular docking, molecular dynamic, cloning, expression and purification of chimeric protein in colorectal cancer treatment. Drug Design, Development and Therapy. 2020 Jan: 309-29. doi: 10.2147/DDDT.S231958.
- [16] Shihab HA, Gough J, Cooper DN, Stenson PD, Barker GL, Edwards KJ, et al. Predicting the functional, molecular, and phenotypic consequences of amino acid substitutions using hidden Markov models. Human Mutation. 2013 Jan; 34(1): 57-65. doi: 10.1002/humu.22225.
- [17] Mahmoud NA, Ahmed DT, Mohammed ZO, Altyeb FA, Mustafa MI, Hassan MA. The Association between SLC25A15 Gene Polymorphisms and Hyperornithinemia-hyperammonemiahomocitrullinuria Syndrome: Using in Silico Analysis. bioRxiv. 2019 Sep: 786301. doi: 10.1101/786301.
- [18] Kumar P, Henikoff S, Ng PC. Predicting the effects of coding non-synonymous variants on protein function using the SIFT algorithm. Nature Protocols. 2009 Jul;

4(7): 1073-81. doi: 10.1038/nprot.2009.86.

- [19] Pires DE, Ascher DB, Blundell TL. mCSM: predicting the effects of mutations in proteins using graphbased signatures. Bioinformatics. 2014 Feb; 30(3): 335-42. doi: 10.1093/bioinformatics/btt691.
- [20] Emadi E, Akhoundi F, Kalantar SM, Emadi-Baygi M. Predicting the most deleterious missense nsSNPs of the protein isoforms of the human HLA-G gene and in silico evaluation of their structural and functional consequences. BMC Genetics. 2020 Dec; 21(1): 1-27. doi: 10.1186/s12863-020-00890-y.
- [21] Ashkenazy H, Abadi S, Martz E, Chay O, Mayrose I, Pupko T, et al. ConSurf 2016: an improved methodology to estimate and visualize evolutionary conservation in macromolecules. Nucleic Acids Research. 2016 Jul; 44(W1): W344-50. doi: 10.1093/ nar/gkw408.
- [22] Alabid T, Kordofani AA, Atalla B, Altayb HN, Fadla AA, Mohamed M, et al. In silico Analysis of Single Nucleotide Polymorphisms (SNPs) in HumanVCAM-1 gene. Journal of Bioinformatics, Genomics and Proteomics. 2016 May; 1(1): 1004.
- [23] Studer G, Tauriello G, Bienert S, Biasini M, Johner N, Schwede T. ProMod3–A versatile homology modellingtoolbox. PLoS Computational Biology. 2021 Jan; 17(1): e1008667. doi: 10.1371/journal.pcbi. 1008667.
- [24] Combet C, Blanchet C, Geourjon C, Deleage G. NPS@: network protein sequence analysis. Trends in Biochemical Sciences. 2000 Mar; 25(3): 147-50. doi: 10.1016/S0968-0004(99)01540-6.
- [25] Lorenz R, Bernhart SH, Höner zu Siederdissen C, Tafer H, Flamm C, Stadler PF, et al. ViennaRNA Package 2.0. 2011; 6: 1- 14. doi: 10.1186/1748-7188-6-26.
- [26] Gao J, Mazor T, Ciftci E, Raman P, Lukasse P, Bahceci I, et al. The cbioportal for cancer genomics: An intuitive open-source platform for exploration, analysis and visualization of cancer genomics data. Cancer Research. 2018 Jul; 78(13_Supplement): 923-. doi: 10.1158/1538-7445.AM2018-923.
- [27] Chen F, Chandrashekar DS, Varambally S, Creighton CJ. Pan-cancer molecular subtypes revealed by mass-spectrometry-based proteomic characterization of more than 500 human cancers. Nature Communications. 2019 Dec; 10(1): 5679. doi: 10.1038/s41467-019-13528-0.
- [28] Tang Z, Li C, Kang B, Gao G, Li C, Zhang Z. GEPIA: a web server for cancer and normal gene expression profiling and interactive analyses. Nucleic Acids Research. 2017 Jul; 45(W1): W98-102. doi: 10.1093/nar/gkx247.

- [29] Zito A, Lualdi M, Granata P, Cocciadiferro D, Novelli A, Alberio T, et al. Gene set enrichment analysis of interaction networks weighted by node centrality. Frontiers in Genetics. 2021 Feb; 12: 577623. doi: 10.3389/fgene.2021.577623.
- [30] Saito R, Smoot ME, Ono K, Ruscheinski J, Wang PL, Lotia S, et al. A travel guide to Cytoscape plugins. Nature Methods. 2012 Nov; 9(11): 1069-76. doi: 10.1038/nmeth.2212.
- [31] Krebs BB and De Mesquita JF. Amyotrophic lateral sclerosis type 20-In Silico analysis and molecular dynamics simulation of hnRNPA1. PloS One. 2016 Jul; 11(7): e0158939. doi: 10.1371/journal.pone.0158939.
- [32] Yazar M and Özbek P. In Silico Tools and Approaches for the prediction of functional and structural effects of single-nucleotide polymorphisms on proteins: an expert review. OMICS: A Journal of Integrative Biology. 2021 Jan; 25(1): 23-37. doi: 10.1089/omi. 2020.0141.
- [33] Baird DM. Variation at the TERT locus and predisposition for cancer. Expert Reviews in Molecular Medicine. 2010 May; 12: e16. doi: 10.1017/S146239941000147X.
- [34] Pak MA, Markhieva KA, Novikova MS, Petrov DS, Vorobyev IS, Maksimova ES, et al. Using AlphaFold to predict the impact of single mutations on protein stability and function. Plos One. 2023 Mar; 18(3): e0282689. doi: 10.1371/journal.pone.0282689.
- [35] Shaw G. Polymorphism and single nucleotide polymorphisms (SNP s). BJU International. 2013 Sep; 112(5): 664-5. doi: 10.1111/bju.12298.
- [36] Fung TS and Liu DX. Post-translational modifications of coronavirus proteins: roles and function. Future virology. 2018 May;13(6): 405-30. doi: 10.2217/fvl-2018-0008.
- [37] Simpson CM, Zhang B, Hornbeck PV, Gnad F. Systematic analysis of the intersection of disease mutations with protein modifications. BMC Medical Genomics. 2019 Jul; 12: 1-0. doi: 10.1186/s12920-019-0543-2.
- [38] Lee DD, Leao R, Komosa M, Gallo M, Zhang CH, Lipman T, et al. DNA hypermethylation within TERT promoter upregulates TERT expression in cancer. The Journal of Clinical Investigation. 2019 Jan; 129(1): 223-9. doi: 10.1172/JCI121303.